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Lobatamide C: Total Synthesis, Stereochemical Assignment, Preparation of Simplified Analogues, and V-ATPase Inhibition Studies

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Abstract: The total synthesis and stereochemical assignment of the potent antitumor macrolide lobatamide C, as well as synthesis of simplified lobatamide analogues, is reported. Cu(I)-mediated enamide formation methodology has been developed to prepare the highly unsaturated enamide side chain of the natural product and analogues. A key fragment coupling employs base-mediated esterification of a β -hydroxy acid and a salicylate cyanomethyl ester. Three additional stereoisomers of lobatamide C have been prepared using related synthetic routes. The stereochemistry at C8, C11, and C15 of lobatamide C was assigned by comparison of stereoisomers and X-ray analysis of a crystalline derivative. Synthetic lobatamide C, stereoisomers, and simplified analogues have been evaluated for inhibition of bovine chromaffin granule membrane V-ATPase. The salicylate phenol, enamide NH, and ortho-substitution of the salicylate ester have been shown to be important for V-ATPase inhibitory activity.

Introduction

Biological metabolites are the subject of ongoing investigations in the search for new medicinal leads including antiinfective and anti-cancer agents. Recently, a number of unique antitumor natural products containing a central benzolactone core bearing an unusual enamide side chain have been reported. Members include salicylihalamides A and B¹ lobatamides A-F,² apicularens A and B,³ CJ-12,950 and CJ-13,357,⁴ and oximidines I and II⁵ (Figure 1). The lobatamides, containing a 15-membered ring macrodilactone and a divinylcarbinol moiety, were isolated in 1998 by Boyd et al. from a southwestern Pacific

tunicate.² Suzumura et al. also independently isolated YM-75518A-C, identical to lobatamides A-C, and the Z-oxime stereoisomer YM-75518D (Figure 1).6 The absolute stereochemistry of the C15 divinylcarbinol of YM-75518A was assigned as (S) using modified Mosher ester analysis.^{6b} However, the configurations of the two remaining stereocenters were not determined and thus required chemical synthesis for full confirmation.

Extensive biological evaluation of lobatamides has been performed against the National Cancer Institute's (NCI) 60 human tumor cell line (mean panel GI₅₀'s approximately 1.6 nM).² Significantly, biological studies indicate that both salicylihalamides and lobatamides represent antitumor natural products with a novel mechanism of action.² Recently, it has been reported that the salicylate enamide macrolides selectively inhibit vacuolar-type proton ATPases (V-ATPases),⁷ ubiquitous proton-translocating pumps of eukaryotic cells.8 Moreover, it has been found that proton-extruding V-ATPases are expressed

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Figure 1. Salicylate enamide natural products.

on the plasma membranes of human tumor cells.9 Accordingly, these natural products are exciting new targets for chemical synthesis, lead optimization studies, and preparation of designed analogues and molecular probes to further define interactions with the molecular target. The salicylihalamides have been synthesized in several laboratories.1c-h De Brabander and coworkers have reported the preparation and structure-function analysis of a number of promising salicylihalamide derivatives.¹⁰ Herein, we report a full account of the total synthesis and stereochemical assignment of lobatamide C,¹¹ as well as structure-activity studies in the lobatamide series.

Development of Methodology for Synthesis of the **Enamide Side Chain**

To enable the synthesis of the salicylate enamides as an entire class, we required a general method to synthesize highly unsaturated enamides. In general, enamides have been shown to have ambident reactivity, both electrophilic at the α -carbon and nucleophilic at the β -carbon. Enamides may be regarded as deactivated enamines and will react with powerful electrophiles such as bromine, peracids, and lead(IV) acetate.¹² They are stable compounds under neutral or basic conditions and with Brønsted acids give rate-determining protonation on carbon, leading to a reactive N-acyliminium ion intermediate that may either undergo hydrolysis of the double bond to form carbonyl compounds and amides13 or react with a range of nucleophiles, including oxygen, sulfur, or π -based nucleophiles.¹⁴

Enamides have been previously synthesized using a number of methods, including N-acylation of imines,¹⁵ elimination of

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α-substituted amides,¹⁶ isomerization of N-allylamides,¹⁷ palladium(II)-catalyzed amidation of alkenes,18 direct addition of amides to alkynes,¹⁹ acid-catalyzed condensation of aldehydes and amides,²⁰ amide Peterson olefination,²¹ Horner-Wittig and Wadsworth-Emmons reactions,^{20b,22} and N-acylation of protected enamines.^{1f,23} Other recent enamide formation methods have also been developed, including organometal addition to vinyl isocyanates,^{5b,24} oxidative decarboxylation-elimination,²⁵ Ru-catalyzed chain extension,²⁶ and rearrangement of N-(α silvl)allyl amides.²⁷ Some of these methods have been employed in the synthesis of enamide-containing natural products such as lycorine,²⁸ mycalolide A,²⁹ chondriamides,²⁵ crocacin D,³⁰ TMC-95 A and B,27 and the salicylihalamides.1c-f

In considering potential new methods for the formation of enamides, we have focused our efforts on transition metalcatalyzed vinylic substitution reactions of vinyl halides and amides, due to the ready availability and stability of the reaction partners and the possibility that such C-N bond constructions may occur in a stereocontrolled manner (Figure 2). It was envisioned that a coupling process could be developed wherein E and Z vinyl halides could be converted to the corresponding

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Figure 2. Stereoselective C-N bond construction for enamide synthesis.

Scheme 1^a



 a Reagents and conditions: (a) CuI (0.10 equiv), Cs₂CO₃ (1.2 equiv), PPh₃ (0.20 equiv), NMP, 90 °C, 12 h, 31% (¹H NMR); (b) CuTC (0.10 equiv), Cs₂CO₃ (1.2 equiv), NMP, 90 °C, 12 h, 59% (¹H NMR).

E and *Z* enamides. This is significant since many of the existing approaches to prepare enamides are not stereoselective. As an entry to our studies, we were attracted to a study by Ogawa et al. who reported the copper iodide-promoted substitution of vinyl bromides and potassium amides (1 equiv of CuI, HMPA, 130 °C) to afford enamides in low to moderate (38–45%) yields.³¹ On the basis of this precedent and related Cu(I)-catalyzed C–N cross coupling reactions,³² we focused on developing a Cu(I)-catalyzed amidation method that would occur at milder temperatures and would be suitable for the installation of potentially labile enamides on complex substrates.³³

Using benzamide and (E)-1-iodo-1-heptene $(1)^{34}$ as model substrates, we initially compared Cu(I) phosphine³⁵ and Cu(I) carboxylate catalysts with Cs₂CO₃ as base (Scheme 1). We obtained a higher conversion (59%) for enamide **2** using Liebeskind's Cu(I) thiophenecarboxylate (CuTC),³⁶ which led us to undertake further optimization with this catalyst.

After reaction optimization, optimal conditions for amidations were discovered by employing CuTC (30 mol %), Cs₂CO₃ as base, and rigorous vacuum purge degassing of the reaction mixture in 1-methyl-2-pyrrolidinone (NMP) prior to heating (90 °C, 12 h). Using these conditions, a number of enamides were prepared as shown in Table 1. Both benzamide and (*E*,*E*)-2,4hexadienamide (**3**) ³⁷ participated in vinylic substitution of **1** to afford enamides **2** and **4** in good yields (entries 1 and 2). It should be mentioned that, under the same conditions using an excess of vinyl iodide **1** (2.2 equiv) and Cs₂CO₃ (2.0 equiv), the coupling reaction with benzamide afforded only enamide **2** (77%) and recovered vinyl iodide **1** (70%) without any evidence of an *N*,*N*-divinyl amide product.³⁸ Using (*E*)- β -iodostyrene

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(5),³⁹ related transformations provided the desired enamides **6** and **7** (entries 3 and 4). The coupling reaction was also successful for secondary amides. Amidations of **1** and **5** with *N*-methylformamide provided enamides **8** and **9** as a mixture of rotamers in good yields (entries 5 and 6). With 2-pyrrolidinone, enamide 10^{20a} was obtained in 99% yield (entry 7). Amidation of a *Z*-vinyl iodide 11^{40} provided *Z*-enamide **12** in only 23% yield (entry 8) likely due to competitive elimination of the *Z*-vinyl iodide to the corresponding terminal alkyne under the reaction conditions.⁴¹

To investigate the nature of the halogen substituent, we evaluated (*E*)-1-bromo-1-heptene $(13)^{42}$ and (3Z)-(4-bromo-3butenyl)benzene⁴³ in coupling reactions with benzamide. However, only trace amounts of enamide products were obtained in these experiments. To compare the reactivities of vinyl iodides and vinyl bromides in the C–N bond formation, we conducted the competition experiment depicted in Scheme 2. Cu-mediated amidation was performed with vinyl bromide **13**, (*E*)-1-iodo-1-pentene (**14**),³⁴ and benzamide in NMP (90 °C). In this case, a 7:1 ratio of enamides **15** and **2** was detected by HPLC⁴⁴ analysis of the crude reaction mixture, which further supports that vinyl bromides are significantly less reactive than vinyl iodides as amidation substrates.⁴⁵

To prepare enamides related to the lobatamides and related salicylate natural products, we next prepared 4-(methoxyimino)-2-butenamides **16** and **17** (Scheme 3). Treatment of 5-hydoxy-2(5H)-furanone⁴⁶ with aqueous methoxyamine hydrochloride led to the formation of 4-(methoxyimino)-(2Z)-butenoic acid **18** (92%).⁴⁷ The corresponding 4-(methoxyimino)-(2Z)-butenamide **16** was prepared in 88% yield by formation of the mixed anhydride of **18** and subsequent reaction with aqueous ammonia. (2Z)-Butenamide **16** could be fully isomerized to (2*E*)-butenamide **17** in 81% yield under acidic conditions.

Next, we conducted model amidation reactions with butenamides **16** and **17** and representative vinyl iodides (Table 2). Treatment of (2*E*)-butenamide **17** with vinyl iodide **1** or **5** using *N*,*N*-dimethylacetamide (DMA) as solvent afforded unsaturated enamides **19** and **20** in 57% and 52% yield, respectively (entries 1 and 2). However, cross-coupling of (2*Z*)-butenamide **16** with vinyl iodide **1** under similar conditions (90 °C, 12 h) did not afford the desired enamide **21**. In an effort to improve this reaction, 2,2,6,6-tetramethyl-3,5-heptanedione⁴⁸ was used as

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Table 1. CuTC-Catalyzed Enamide Coupling of Amides and Vinyl Iodides^a



^{*a*} Performed using 30 mol % of CuTC, 1.0 equiv of vinyl halide, 1.5 equiv of amide, and 1.5 equiv of Cs_2CO_3 , NMP, 90 °C, 12 h. ^{*b*} Isolated yields after silica gel chromatography. ^{*c*} 3:1 mixture of rotamers by NMR (400 MHz). ^{*d*} 1.3:1 mixture of rotamers by NMR (400 MHz). ^{*e*} Reaction performed at 60 °C.

Scheme 2. Vinyl Bromide vs Iodide Competition Experiment



ligand to increase the reactivity and turnover of CuTC (Scheme 4). However, the reaction afforded only 20% of enamide **21** along with the unexpected *N*,*N*-divinyl amide **23** (17%).³⁸ Finally, enamide **21** was obtained in 52% yield using Rb₂CO₃ as base⁴⁹ and *N*,*N*'-dimethylethylenediamine (**24**) as ligand^{32d,e} (entry 3). Entry 4 illustrates the stereoselective coupling of *Z*-amide **16** and *Z*-vinyl iodide **11**. The reaction provided a modest yield (30%) of the desired *Z*-enamide **22** when only CuTC was used as catalyst. However, when diamine ligand **24**

Scheme 3



was added, the yield of enamide **22** was enhanced to 55%. This initial study provided encouragement for the stereospecific construction of the *Z*-enamide side chain of oximidines (cf. Figure 1).

Since diamine ligand 24 remarkably facilitated the production of enamides 21 and 22, we employed 24 as ligand in couplings to prepare enamides 19 and 2. However, the yield of 19 (54%) was slightly lower than that solely using CuTC as catalyst (57%), while the yield of 2 was only 40%.⁵⁰ These results

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Table 2. Cu(I)-Catalyzed Vinylic Substitution of Butenamides ${\bf 16}$ and ${\bf 17}$

entry	amides	vinyl iodide	enamide	yields ^a
1	17	1	MeO ⁻ ^N H 19	57 ^b
2	17	5	MeO ^{-N}	52 ^b
3	16	1	MeO ^{-N} O ^{-N} H 21	52 ^c
4	16	11	MeO ^N ONH 22	55 ^d

^{*a*} Isolated yields after silica gel chromatography. ^{*b*} Reaction performed with 0.3 equiv of CuTC, 1.5 equiv of **17**, 1.5 equiv of Cs₂CO₃, DMA, 90 °C, 12 h. ^{*c*} 0.5 equiv of CuTC, 0.5 equiv of **24**, 1.5 equiv of **16**, 1.1 equiv of Rb₂CO₃, DMA, 60 °C, 12 h. ^{*d*} 0.3 equiv of CuTC, 0.6 equiv of **24**, 1.5 equiv of **16**, 1.1 equiv of Rb₂CO₃, DMA, 60 °C, 14.5 h.

Scheme 4



indicate that ligand 24 is most effective when used with (2Z)butenamide 16. It is conceivable that enamide 21 may chelate with CuTC to form complex 25 (Scheme 5), which may substantially deactivate the catalyst. Addition of diamine 24 may compete with enamide 21 to transform a complex such as 25 to 26, which may be available for further catalysis.

First Retrosynthetic Analysis of Lobatamide C

After the establishment of a workable methodology for the synthesis of the enamide side chain, we initiated studies toward the total synthesis of the lobatamides. Our plan for the synthesis of our first target, lobatamide C (**27**), is outlined in Figure 3. In contrast to previous syntheses of the salicylate enamide natural products^{1c-g,3c,d} in which the enamide side chain was typically installed at a late stage in the synthesis, our general plan for lobatamide C was to pursue a convergent coupling of fragments with the enamide side chain preinstalled. This synthetic plan was primarily based on our general concern that base-catalyzed



amidation to construct enamides may be difficult on fragile and potentially elimination prone macrodilactone substrates. As we were later to discover, this convergent approach creates synthetic challenges with regard to manipulation and reactivity of enamide-containing intermediates but also adds an element of flexibility for construction of analogues that vary at the salicylate portion.

Retrosynthetic analysis of the lobatamide C skeleton reveals two principal fragments: the C1–C10 enamide sector **28** and the C11–C26 salicylate subunit **29** containing a divinylcarbinol moiety and an acetonide protecting group. We planned to prepare the precursor **30** from esterification of subunits **28** and **29**, which could be desilylated and macrocyclized by removal of the 1,3-dioxin-4-one to furnish the salicylate ester under basic⁵¹ or potentially photochemical conditions.⁵² Although the configuration of lobatamide C at C8 and C11 was not determined,² we first focused on preparation of the 8*S* enantiomer of **30** in light of the stereochemistry of salicylihalamides^{1c} and



Figure 3. First retrosynthetic analysis for lobatamide C.

⁽⁵⁰⁾ Conditions for preparation of **19**: 30 mol % of CuTC, 60 mol % of **24**, 1.0 equiv of vinyl iodide **1**, 1.5 equiv of amide **17**, 1.1 equiv of Cs₂CO₃, DMA, 60 °C, 12 h. For preparation of **2**: 30 mol % of CuTC, 60 mol % of **24**, 1.0 equiv of vinyl iodide **1**, 1.5 equiv of benzamide, 1.5 equiv of Cs₂CO₃, NMP, 90 °C, 12 h.



oximidines⁵ (cf. Figure 1). The stereochemistry at C8 could easily be altered in the event that this prediction was found to be incorrect. Further disconnection of fragment 29 at the C18-C19 and C14-C15 bonds led to benzyl bromide 31, Z-vinyl stannane **32**,⁵³ and nonracemic alkyne **33**. The divinylcarbinol moiety may be obtained by addition of a vinylzinc species (derived from 33 by hydrozirconation and transmetalation),⁵⁴ to an enal which will be prepared by Stille coupling⁵⁵ of 31 and 32, desilylation, and alcohol oxidation. It was anticipated that stereocontrol at C15 could be established using chiral ligands in the vinylzinc addition step.56 Enamide subunit 28 may be prepared by Cu(I)-catalyzed amidation of vinyl iodide 34 with amide 17 according to the C-N bond formation protocols previously described.

Synthesis of C1–C10 Subunit (28)

Preparation of a protected form of the C1-C10 enamide fragment 28 is shown in Scheme 6. Ethyl vinyl acetate was transformed in quantitative yield into (\pm) -ethyl-3,4-epoxybutanoate 35 using m-CPBA.⁵⁷ Hydrolytic kinetic resolution (HKR) developed by Jacobsen and co-workers⁵⁸ was used to

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(57)

establish the C8 stereocenter and efficiently prepare multigram amounts of (R)-35 (>99% ee). (R)-35 was next employed in epoxide ring-opening reactions in order to prepare a vinyl iodide substrate for enamide synthesis. Addition of the acetylenic alane reagent⁵⁹ derived from trimethylsilylacetylene led to facile epoxide opening and afforded 36 in 80% yield. Silvl protection of 36 using chlorodiethylisopropylsilane (DEIPSCI), followed by treatment of the resulting silyl ether 37 with AgNO₃/NBS/ H₂O,⁶⁰ afforded the bromoalkyne intermediate **38** that was converted to the (E)-stannylalkene **39** using Pattenden's method⁶¹ (78%, three steps). Iodine exchange of 39 furnished vinyl iodide **34** in 73% yield with full retention of olefin stereochemistry. In contrast, iodination of the desilvlated analogue of **39** afforded a 5:1 E:Z mixture of vinyl iodides, indicating the importance of protecting the secondary alcohol. After considerable experimentation, we found that Cu(I)-mediated vinylic substitution of **34** with the butenamide **17** (CuTC, 1,10-phenanthroline,^{32a} dba, Cs₂CO₃, DMA) led to a 52% yield of the C1-C10 enamide fragment 40. Critical to the success and reproducibility of this reaction were the use of a moderate reaction temperature (65

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Scheme 8



Scheme 10



°C) to suppress elimination of the sensitive β -silvloxy ester coupling substrate, the use of 1,10-phenanthroline as supporting ligand for CuTC, and use of high purity cesium carbonate.⁶² Synthesis of the target C1-C10 fragment was completed by hydrolysis of 40 to afford the labile enamide acid 28 (73%).

with preparation of salicylate benzyl bromide 31, prepared in two steps from the known aryl triflate 41^{63} (Scheme 7). Treatment of **41** with lithium dimethylcuprate⁶⁴ and MeI⁶⁵ led to the production of 6-methylsalicylate derivative 42 that was brominated⁶⁶ to afford benzylic bromide **31** (78%). Coupling of **31** with vinylstannane 32^{53} using the conditions reported by Kamlage⁶⁷ (Pd₂dba₃-CHCl₃, AsPh₃, THF) afforded Z-allylic silvl ether 43 (79%). Silvl deprotection using HF/pyridine provided allylic alcohol 44, which was oxidized to Z-enal 45 using Dess-Martin periodinane.⁶⁸ However, **45** was found to be readily isomerized to E-enal 46 during purification on silica gel (46:45 = 6:1). Hence, crude 45 was used in subsequent coupling reactions. Hydrozirconation of DEIPS-protected (R)-3-butyn-2-ol (33), followed by transmetalation with Et_2Zn and addition to 45 according to the general procedure of Wipf and

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⁽⁶⁵⁾ Quenching the reaction with MeI was found to significantly improve the reaction yield, likely due to methylation of an aryl copper side product. Cf. Cohen, T.; Wood, J.; Dietz, A. G., Jr. Tetrahedron Lett. 1974, 15, 3555.



Figure 4. Proposed transesterification using intramolecular general base catalysis.

Scheme 13



co-workers,⁵⁴ led to the target divinylcarbinol fragment 47 as a 1:1 mixture of diastereomers (30% yield). Although the Z-olefin stereochemistry of 47 was verified using NOE experiments, the presence of significant amounts of isomerized E-enal 46 led to rapid alteration of the synthetic sequence and use of configurationally stable Z-enal 48⁶⁹(Scheme 8). In the event, hydrozirconation, zirconocene-zinc transmetalation and addition to enal **48** afforded divinylcarbinol **49** as a 1:1 nonseparable mixture of diastereomers (87%). Compound 49 was further advanced by silvlation of the secondary alcohol, followed by lithiationtrimethylstannylation, to afford vinyl stannane 51 which was coupled with benzyl bromide 31 to furnish C11-C26 fragment 52 (56%). Selective deprotection of 52 using HF/pyridine at 0 °C afforded the target alcohol 29 (69%).

Fragment Coupling and Attempted Macrocyclization

With fragments 28 and 29 in hand, we attempted their coupling using standard acylation methods. However, both Yamaguchi⁷⁰ and Keck⁷¹ esterification conditions did not furnish the desired ester C11-epi-30, presumably due to decomposition of the sensitive enamide side chain. However, the esterification was successfully accomplished using Mitsunobu conditions to afford 30 (70%, Scheme 9). Selective deprotection of the C8 DEIPS ether of 30 using HF/pyridine afforded acyclic alcohol 53 (41%). Extensive studies were performed to prepare macrolactone 54 by intramolecular transesterification of 53. Treatment of 53 under basic conditions (NaH51 or NaHMDS, THF) or distannoxane transesterification catalysts⁷² did not furnish the desired cyclized product 54. Under these conditions,



Figure 5. Revised retrosynthesis of lobatamide C

Scheme 14





Scheme 15



compound 53 was substantially decomposed. Attempted photolysis of 53 (Ace-Hanovia UV lamp 450 W, Pyrex filter, 20 min) in an effort to form the macrolactone by trapping of a keto-ketene intermediate52 led to complete decomposition of the substrate. Due to a concern that the C15 TBS ether may block macrocyclization, we next prepared compound 55 by removing the TBS ether with HF/pyridine. However, treatment of 55 under basic conditions (NaH, THF) led to the production of the eight-membered lactone enamide 56 (dr 1:1) instead of the desired natural product. Compound 56 is formally an isomer of lobatamide C in which the divinylcarbinol forms a salicylate ester, leaving a pendant C8 hydroxyl. Attempts to rearrange 56 to 15-membered ring lobatamide C (27) by translactonization

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Scheme 16



1) Bu₄N⁺OH⁻

of water

80 °C, 2 h

Scheme 17



protocols73 (e.g., catalytic PPTS/CH2Cl2 or Et2Zn/ CH2Cl2) were not successful.

Development of Methodology for the Construction of Salicylate Esters

In parallel with our total synthesis efforts, we initiated studies toward the synthesis and biological evaluation of simplified analogues of the lobatamides in order to clarify the minimal core structure (pharmacophore) required for V-ATPase inhibition. In initial studies, a series of acyclic analogues were prepared using the C1-C10 enamide fragment. Our first target was simplified salicylate enamide 57 that eliminates the C8 stereogenic center (Scheme 10). TIPS-protected vinyl iodide 5874 underwent CuTC-mediated cross-coupling with (2E)-butenamide 17 to furnish enamide 59, which was desilylated using TBAF to afford enamide alcohol 60. Encouraged by patent literature for esterification of salicylate cyanomethyl esters⁷⁵ and general stability of enamides to basic conditions, we first treated enamide alcohol 60 with cyanomethyl ester 61^{75} in the presence of a catalytic amount of K2CO3 (DMA, 90 °C). Salicylate enamide 57 was rapidly and cleanly produced under these conditions (91%).

As a control experiment, we found that the acylation using benzovloxyacetonitrile 62 with alcohol 60 under similar reaction conditions did not afford the desired ester product (Scheme 11). We initially suspected that keto-ketene 63 was the active acylating agent.⁷⁶ However, trapping experiments with N,N-



dimethylcyanamide, ethyl vinyl ether, ethoxyacetylene, and N-benzylmaleimide failed to provide the corresponding Diels-Alder adducts.⁷⁷ Interestingly, treatment of **60** and methoxy cyanomethyl ester 64 under the same conditions afforded the ester product 65, albeit in lower chemical yield (28%) and limited conversion (Scheme 11). These results, in conjunction with conformational analysis of the sodium salt of 61,78 suggested an alternative mechanistic pathway involving intramolecular general base catalysis. In the conformer shown in Figure 4, the ester carbonyl is out of planarity, which is expected to increase the reactivity of the carbonyl toward transesterification. In addition, the o-hydroxyl may be oriented to act as a general base catalyst and direct attack of the alcohol to the π^* of the salicylate carbonyl. This proposed transesterification mechanism is substantiated by literature precedent.⁷⁹

To ultimately apply this transesterification methodology to the construction of the salicylate portion of lobatamide C, we next prepared analogues 66-68. In particular, compounds 66 and 68 contain o-salicylate alkyl substituents. Hydrolysis of benzodioxinone 42 with KOH quantitively provided 6-methylsalicylic acid⁸⁰ (Scheme 12), which was converted to cyanomethyl ester 69 (80%).⁸¹ Acylation of enamide alcohol 60 with 69 in the presence of K_2CO_3 in DMA afforded analogue 66 (70%). Analogue 67 was prepared from secondary alcohol 70 (prepared by TBAF desilylation of 40), which exhibited significantly lower reactivity in acylation with cyanomethyl ester 61 (25% yield, Scheme 13). In addition, 15% of the β -elimination product, as well as considerable amounts of recovered 70, were obtained. Under similar conditions, acylation of 70 with cyanomethyl ester 69 did not afford the desired salicylate product. However, the derived acid **71**, after neutralization with Bu₄NOH and heating with 69 and Na₂CO₃ (1.0 equiv) in DMA/ 2-butanone (80 °C, 2 h), furnished the desired salicylate ester. The tetrabutylammonium salt of **71** both increases the solubility of the enamide alcohol fragment⁸² and likely blocks α-deprotonation/elimination of the β -salicyloxyester product. A number

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of carbonate bases (Li, K, Rb, and Cs) were also evaluated, but interestingly only stoichiometric levels of Na₂CO₃ were effective. To facilitate purification, the unstable enamide acid product was methylated with trimethylsilyl diazomethane to afford methyl ester **68** (34%, three steps).

Revised Retrosynthesis of Lobatamide C

Due to the difficulties in forming the C8 salicylate ester using macrocyclization (cf. Scheme 9), we altered our synthetic route by constructing this bond at an earlier stage using the cyanomethyl ester transesterification methodology successfully employed in our simplified analogue synthesis. Figure 5 illustrates our revised retrosynthetic analysis of lobatamide C (27) by macrolactonization of hydroxy acid 72 to form the C10–C11 ester bond and deprotection of the silyl group at the C15 divinylcarbinol. Acid 72 may be prepared by base-catalyzed transesterification of cyanomethyl ester fragment 73 and hydroxy enamide acid 71. Further, disconnection of fragment 73 at C18–C19 bond reveals benzylic bromide and vinylstannane fragments which may be merged by sp²–sp³ coupling (cf. Scheme 8).

Completion of the Total Synthesis of Lobatamide C

Synthesis of fragment **73** requires diastereomerically pure divinylcarbinol **49**. Compound **49** (dr 1:1) was prepared from alkyne **33** (Scheme 8). To establish the proposed *S* configuration of the divinylcarbinol at C15, extensive studies were performed, including evaluation of numerous amino alcohol controllers.⁸³ However, the best result (2:1, 15S:15R) was obtained using Wipf's amino thiol ligand **74**⁵⁴ (Scheme 14). Compound **49a** (2:1 dr) was next advanced to vinyl stannane **51a**. Silyl protection of cyanomethyl ester **69**, followed by benzylic bromination, afforded benzylic bromide **76** (Scheme 15). Stille coupling of **76** and vinyl stannane **51a** afforded C11–C26 fragment **77**. Selective desilylation of **77** (TBAF, 0 °C) afforded the target salicylate cyanomethyl ester fragment **73**.

After preparation of fragments **71** and **73**, we found that the tetrabutylammonium salt of enamide acid **71** participated in smooth esterification reactions with cyanomethyl ester **73** (Na₂CO₃, DMF/2-butanone, 80 °C, 2 h) to provide the desired salicylate **78** (Scheme 16). Treatment of **78** with HF/pyridine (rt, 1 h) afforded seco acid **72** (43%, two steps). Both **78** and **72** are highly labile compounds and could only be purified using





reverse phase (C18) silica. This instability is likely due to the pendant carboxylic acid that may protonate and decompose the enamide functionality.¹³ Interestingly, when acid **78** was exposed to HF/pyridine for extended reaction times (4 h), the eight-membered ring lactone **79** was obtained along with desired compound **72** in modest yield (Scheme 17). To further confirm the structure, compound **79** was coupled with enamide acid **28** under Mitsunobu conditions and desilylated to **56** (50%, 2 steps), which was obtained in our earlier synthetic route (cf. Scheme 9).

Gratifyingly, seco acid **72** was smoothly macrolactonized using intramolecular Mitsunobu conditions⁸⁴ to afford separable macrolactones **54a** (26%) and **54b** (26%) (Scheme 18). However, the formation of **54a** and **54b** in a 1:1 ratio indicates influence of the protected divinylcarbinol stereocenter on the macrocyclization and thus necessitated independent confirmation of the C15 stereochemistry. Desilylation of **54a** and **54b** with HF–pyridine/pyridine led to efficient production of lobatamide C (**27a**) (52%) and its C15 epimer **27b** (78%). Synthetic **27a** was confirmed to be identical to data reported for natural lobatamide C by ¹H and ¹³C NMR, $[\alpha]_D$ (–18.8°, *c*

⁽⁸³⁾ In addition to chiral ligand-controlled vinyl zinc addition to aldehydes, other methods were also evaluated to establish the C15 (S) configuration, including lipase resolution, asymmetric reduction of the corresponding divinyl ketone, and zinc triflate-mediated asymmetric alkynylation of enal 48 (cf. Frantz, D. E.; Fässler, R.; Carreira, E. M. J. Am. Chem. Soc. 2000, 122, 1806). However, none of these methods afforded useful levels of selectivity.

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Figure 6. X-ray Crystal Structure of 84.

0.17, MeOH), TLC R_f values in three solvent systems, and COMPARE profiles in NCI's 60 tumor cell line.⁸⁵

Stereochemical Assignment of Lobatamide C (27a)

The absolute configuration of 27a at C15 was determined to be S using a modified Mosher's ester analysis according to Suzumura's procedure.⁸⁶ In addition, 11R diastereomers of lobatamide C (27c and 27d) were prepared independently from (S)-3-butyn-2-ol (Scheme 19). TIPS-protected (S)-3-butyn-2ol 80 was converted to divinylcarbinol 81 (dr = 1:1) using hydrozirconation-transmetalation-enal addition. Fortunately, diol 82 (dr = 1:1, obtained by TBAF desilylation of 81) was resolved by recrystallization in CHCl₃. Using this method, diastereomers 82a and 82b were obtained in high diastereomeric purity (>95% de).⁸⁷ Selective protection of diols 82a and 82b with DEIPSCI furnished divinylcarbinols 83a and 83b. Lobatamide C diastereomers 27c and 27d were synthesized from 83a and 83b, respectively, employing the same route. Both HPLC and NMR studies indicated that 27c and 27d did not match natural lobatamide C.

Although numerous allylic alcohols have been reported to undergo inversion in Mitsunobu reactions,⁸⁸ recent work has documented examples of retention of stereochemistry in Mitsunobu esterifications.⁸⁹ To fully confirm the stereochemical assignment of lobatamide C, compound **54a** was derivatized to afford bromophenyl acetate **84** (Figure 6). Single X-ray crystal structure of **84** indicates that the stereochemistry at C11 is *S*, clearly showing that the C11 stereogenic center was inverted in the Mitsunobu macrocyclization. The crystal structure of **84** thus fully confirms the stereochemistry of lobatamide C as 8*S*, 11*S* and 15*S* without ambiguity.

Synthesis and Biological Evaluation of Simplified Lobatamide Analogues

Although several simplified analogues (57, 65-68) were prepared during the fragment coupling studies, we synthesized an additional lobatamide derivative 85 bearing an *N*-methyl en-



Figure 7.

amide and a macrocyclic analogue **86**, which replaces the fragile divinylcarbinol moiety of lobatamide C with a Z-olefin and a simplified three-carbon segment (Figure 7).⁹⁰ The preparation of **86** is shown in Scheme 20. Stille coupling of vinyl stannane **87**⁹¹ and benzylic bromide **76** afforded salicylate **88**. Selective desilylation (TBAF) provided cyanomethyl ester **89**, which was coupled with enamide acid **71** to furnish the salicylate **90**. Desilylation (HF–pyridine/pyridine) provided hydroxy acid **91** which underwent Mitsunobu macrolactonization (0.005 M) to afford 14-membered macrolactone analogue **86** (65%).

Synthetic lobatamide C (27a), stereochemical isomers (27b-d), and lobatamide analogues were evaluated for activity against bovine V-ATPase from chromaffin granule membranes. As shown in Table 3, both lobatamide C and its C15 epimer **27b** showed potent inhibition (2.1 and 3.6 nM), while the 11Risomers of lobatamide C (27c.d) showed lower activity (20 and 21 nM). The eight-membered ring lobatamide C isomer 56 had significantly reduced activity. For simplified lobatamide analogues, compounds 57 were found to inhibit bovine V-ATPase with modest activity. Methylated analogues 65 and 85 had significantly reduced activities against V-ATPase, which indicates the importance of a free phenol and enamide NH. Macrodilactone **86** showed good V-ATPase inhibition $(1.4 \,\mu\text{m})$, but not at the nanomolar potency of the lobatamides, which indicates that the ring size and substitution of the macrolactone are critical for potent V-ATPase inhibition. Interestingly, permutation of the *ortho* hydrogen of analogue 57 to a methyl group substantially increased the activity (66, 1.6 μ M). 6-Methyl salicylate compound 68 also showed potent inhibition of V-ATPase (0.1 μ M), approximately 180 times more active than compound 67 without a methyl group ortho to the carbonyl. Conformational analysis⁷⁸ of **67** and **68** (Figure 8) indicates that conformers of ortho-H substituted salicylate 67 maintain near planarity of the carbonyl with the aromatic ring. In contrast, in analogue 68 the o-methyl substituent forces the carbonyl outof-plane by approximately 60°, presumably because of steric

⁽⁸⁵⁾ Synthetic 27a and 27b were evaluated against NCI's 60 tumor cell line. Mean panel GI₅₀'s: 3.7 nM (natural lobatamide C), 5.1 nM (27a), and 227 nM (27b); GI₅₀ (COMPARE correlation): 1.00 (natural lobatamide C), 0.83 (27a), 0.80 (27b).

⁽⁸⁶⁾ See Supporting Information. We thank Dr. K. Suzumura (Yamanouchi Pharmaceutical Co.) for providing experimental details on the Mosher ester analysis of YM75518A.

⁽⁸⁷⁾ The absolute configurations of diols 82a and 82b were assigned by HPLC analysis and comparison of optical rotations of diols obtained from compound 49. See Supporting Information.

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 Smith, A. B., III.; Safonov, I. G.; Corbett, R. M. J. Am. Chem. Soc., 2002, 124, 11102. (c) Liao, X.; Wu, Y.; De Brabander, J. K. Angew. Chem., Int. Ed. 2003, 42, 1648.

⁽⁹⁰⁾ For a preliminary account, see: Shen, R.; Lin, C. T.; Bowman, E. J.; Bowman, B. J.; Porco, J. A., Jr. Org. Lett. 2002, 4, 3103.







Table 3. Effect of Simplified Analogues of the Lobatamidesagainst Bovine V-ATPase^a

compd	IC ₅₀ (µM)	compd	IC ₅₀ (µM)
27a	0.0021	66	1.6
27b	0.0036	67	18
27c	0.020	68	0.10
27d	0.021	85	>200
56	b	86	1.4
57	с	92	0.21
65	no effect	93	0.06

^{*a*} ATPase activity determined as described in the Supporting Information using 20 μ g of membrane protein and the indicated amount of inhibitor. ^{*b*} 27% inhibition at 30 μ m, higher concentrations not soluble. ^{*c*} 25% inhibition at 20 μ m, higher concentrations not soluble.



Figure 8. Representative minimum energy conformations of **67** (A) and **68** (B) (Chem 3D, enamide side chain omitted for clarity).

hindrance effects.⁹² This steric effect is also seen in the X-ray crystal structure of apicularen A^{3b} and compound **84** (salicylate ester carbonyl out of planarity by 80° and 52°, respectively) and may be an important conformation of the salicylate moiety in potent V-ATPase inhibitors.

On the basis of this observation, compound **92** and **93** with sterically demanding substituents (iodine and prenyl) *ortho* to the carbonyl were prepared in order to potentially increase V-ATPase inhibition potency. Na₂CO₃-mediated coupling of enamide acid **71** and iodo cyanomethyl ester **94** (prepared from 6-iodosalicylic acid⁹³) afforded analogue **92** after methylation (Scheme 21). Preparation of prenylated analogue **93** commenced with triflate **41** (Scheme 22). Stille coupling of **41** and prenyl stannane **95** afforded acetonide **96**, which was hydrolyzed and reprotected as cyanomethyl ester **98**. Analogue **93** was obtained using similar transesterification conditions. Evaluation of **92** and **93** against bovine V-ATPase showed that **92** has lower inhibition activity (210 nM) than analogue **68**, while **93** showed slightly higher activity (60 nM). These results indicate that it is possible to produce relatively potent, acyclic salicylate enamide







Scheme 22



V-ATPase inhibitors. However, in addition to steric hindrance effects, specific orientation of the functional groups by the macrolactone scaffold may be necessary to achieve low nano-molar inhibition against mammalian V-ATPases.

Conclusion

We have achieved a highly convergent synthesis of the potent antitumor and V-ATPase inhibitory natural product lobatamide C. For construction of the enamide side chain of lobatamides and related natural products, we have developed a mild and stereoselective Cu(I)-catalyzed vinylic substitution protocol which complements related C–N bond formation methodolo-

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gies.³² A key step in the total synthesis involves base-mediated esterification of hydroxy enamide and salicylate cyanomethyl fragments. Macrocyclization was achieved using an intramolecular Mitsunobu reaction. The stereochemistry of lobatamide C was assigned as 8S,11S,15S by comparison to that of synthesized diastereoisomers. Preparation and X-ray crystallographic analysis of a lobatamide C derivative fully confirmed the stereochemical assignment. A number of simplified lobatamide analogues have been prepared in an effort to probe structure-activity relationships. Lobatamide C isomers and simplified analogues were evaluated against bovine V-ATPase from chromaffin granule membranes, which showed that the salicylate phenol, enamide NH, and ortho-substitution of the salicylate ester are important for V-ATPase inhibition. In addition, a number of nanomolar acyclic salicylate enamides inhibitors of bovine V-ATPase were uncovered in this study. Further studies on the application of Cu(I)-catalyzed vinylic substitution and preparation of other lobatamide derivatives are in progress and will be reported in due course.

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Supporting Information Available: Experimental procedures and characterization data for all new compounds, including X-ray crystal structure coordinates for **84**. X-ray crystallographic file in CIF format. This material is available free of charge via the Internet at http://pubs.acs.org

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